

Livestock Feed Production from Sago Solid Waste by Pretreatment and Anaerobic Fermentation Process

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Abstract. Food needs in Indonesia is increasing, including beef. Today, Indonesia has problem to do self-sufficiency in beef. The cause of the problem is the quality of local beef is still lower compared with imported beef due to the quality of livestock feed consumed. To increase the quality of livestock is through pretreatment and fermentation. Source of livestock feed that processed is solid sago waste (*Arenga microcarpa*), because in Indonesia that is relatively abundant and not used optimally. Chemical pretreatment process for delignification is by using NaOH solution. The purposes of this research are to study NaOH pretreatment, the addition of *Trichoderma sp.*, and fermentation time to improve the quality of sago solid waste as livestock feed through anaerobic fermentation. The variables used are addition or without addition (4%w NaOH solution and *Trichoderma sp* 1%w) and fermentation time (7, 14 and 21 days), with the response of crude fiber and protein. The result of this research shows that the pretreatment with soaking of NaOH solution, addition of *Trichoderma sp* and 14 days of fermentation was more effective to improve the quality of solid sago waste with decrease of crude fiber from 33.37% to 17.36% and increase of crude protein from 4.00% to 7.96%.

1 Introduction

The Government of Indonesia has made a solution to continue to reduce the famine by making food self-sufficiency policy. Beef is one example of the food. Beef is a kind of food that has multiple roles, both in terms of food security and economy. The increase in beef prices is currently the result of an imbalance between production quotas and the level of public demand for beef [1]. The consequence is that Indonesia must import beef. Based on statistical data on import and export volume of beef during 1996-2015 [2], the import volume of beef in Indonesia tends to increase every year with the highest number obtained in 2014 of 246,000 tons. The data shows that Indonesia is still can't to be self-sufficient in beef. The imported beef in Indonesia comes from the United States has a smooth and soft texture, contains high protein and beneficial to the human immune. This meat also has fat (marbling), as other premium beef meat, such Angus beef. The quality of USA beef is different when compared to the quality of local beef because of differences in the livestock feed [3].

Indonesia's conventional livestock feed products are declining due to weather conditions and fluctuations in price. This reason is causing for seeking alternative sources, environmentally friendly, economically acceptable, and socially acceptable non-conventional feed sources [4]. The livestock feed should also be able

to become a mainstay feed in the long term. Potential and useful alternative livestock feed and also available throughout the year are generally from agricultural processing industries, for example, in the production of sago flour [5]. Sago plants (*Arenga Microcarpa*) grow in damp tropical regions. Currently, the use of sago only focuses on the starch contained in it [6]. The processing of sago flour is usually conducted near water sources such as rivers or creeks edge [7]. The process of sago flour processing in the palm sugar industry in Daleman village, Klaten regency, Central Java produces an average of 200 tons / year of sago flour with 659 tons / year or 2.19 tons / day [8].

Solid sago waste contains lignocellulose rich in cellulose that can be used optimally as a carbon source. The crude protein content of the sago waste is very low, i.e. 2.63 %, while the crude fiber content is 15.90 % [9]. Sanchez [10] states that Cellulose and hemicellulose are also substances of plant tissue composed of different sugars. Cellulose is the main material that encloses the plant skeleton resulting from the process of photosynthesis of plants. This compound is not soluble in ordinary solvents and comprises a plurality of β -D-glucopyranose units connected via β -1-4 bonds to form straight and long chains reinforced by a cross-linked hydrogen bond providing a crystalline structure and arranged in a micro fibril. Hemicellulose is the second largest polysaccharide after cellulose. Hemicellulose is a complex polymer consisting of a mixture of different

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monosaccharide polymers in which the constituent sugars are grouped in the form of hexoses (glucose, mannose, and galactose), pentose (silose, arabinopiranos, arabinofuranosa), hexuronic acid (glucuronate, methylglucoromate, galactoromat), deoxyhexose (rammosa, fucosa) [11].

Lignin is an anti-nutrient for livestock that interfered with livestock digestion and decrease the nutritional value of animal feed ingredients [12]. Lignin is an undigested component, affecting the digestibility of crude fiber. This compound is an aromatic polymer of phenylpropanoid produced from synthesis of coniferyl, synaphyl, p-coumaryl alcohol. Lignin is highly resistant to any chemical degradation including enzymatic. Bases pretreatment usually use basic solutions such as NaOH, Ca(OH)₂ (limestone) or ammonia to remove lignin and parts of hemicellulose, so this process can efficiently increase cellulosic enzyme capability [13]. Generally this pretreatment is conducted under low temperature conditions with long time and high base concentrations [14]. The advantage of using a base as a lignocellulose pretreatment is the presence of lignin destruction wherein lignin inhibits the action of the enzyme during the anaerobic process [15]. Alkali can damage ester bonds between lignin and xylem [16].

Fermentation is one of the ways to improve the quality of the material where in the process there is an overhaul of hard structures physically, chemically and biologically so that the material from complex structures becomes simple, thereby making the digestibility more efficient [17]. According to Oseni and Ekperigin [18], Fermentation of plant waste not only increases the contents of protein, fat or crude fiber, but also increases the content of essential minerals that can be useful to the body's metabolism. The purposes of this research are to study NaOH pretreatment, the addition of *Trichoderma* sp and fermentation time to improve the quality of solid sago waste as livestock feed through anaerobic fermentation.

2 Materials and Method

2.1 Materials

The material used in this study is solid sago waste from Jepara District (Central Java), *Trichoderma* sp from Pradipta Paramita Solo, molasses from Ungaran, NaOH, aquadest, and acetic acid.

2.2 Preparation Stage

At this stage, the sago palm waste raw material is collected from sago palm processing industry in Plajan village, Jepara District, Central Java Province. *Trichoderma* sp was purchased from Pradipta Paramita Solo, molasses from Ungaran, while NaOH and acetic acid from the Indrasari chemical store. After that, preparation is conducted by sieving the solid sago waste and each variable is measured 50 grams.

2.3 Operation Stage

Solid sago waste was soaked with 4% w NaOH solution for 24 hours or heated with 4% w NaOH solution at 60OC for an hour. The pretreatment substrate is reduced to a pH of 5-6 with acetic acid and then fermented for 7, 14 and 21 days with *Trichoderma* sp and molasses. Fermentation process in this study conducted anaerobically.

2.4 Result Analysis Stage

The analysis was conducted graphically based on the phenomenon for the effect of the independent variables on the contents of crude fiber and crude protein.

3 Results and Discussion

3.1 Effect of pretreatment type on Crude Fiber and Crude Protein Content

Figure 1 and 2 shows the correlation of the pretreatment of NaOH and the crude fiber content of the feed material derived from the solid sago waste. In Figure 1, the solid sago waste variables without the addition of *Trichoderma* sp on the day 7 samples without pretreatment, heating pretreatment, and soaking pretreatment resulted in crude fiber content of 33.04%; 27.50% and 22.03%, respectively, on the day 14 samples without pretreatment, heating pretreatment, and soaking pretreatment resulted in crude fiber content of 33.01%; 27.49% and 22.63%, respectively, on the day 21 samples without pretreatment, heating pretreatment, and soaking pretreatment resulting in crude fiber content respectively 33.87%; 27.98% and 23.00% respectively. In Figure 2, the solid sago waste variables with the addition of *Trichoderma* sp on the day 7 samples without pretreatment, heating pretreatment, and soaking pretreatment resulted in crude fiber content of 30.00%; 27.01% and 21.79%, respectively, on the day 14 samples without pretreatment, pretreatment heating and soaking pretreatment resulted in crude fiber content 25.18%; 21.71% and 17.37%, respectively, on the day 21 samples without pretreatment, heating pretreatment, and soaking pretreatment resulted in crude fiber content of 27.62%; 24.53%, and 19.11%, respectively.

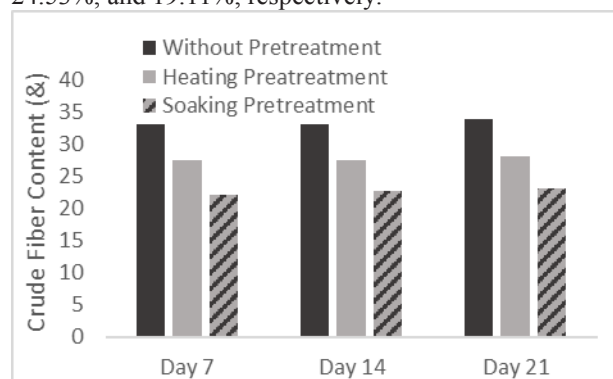


Fig. 1. Correlation of NaOH Pretreatment vs. Crude Fiber Content in Variables without Addition of *Trichoderma* sp.

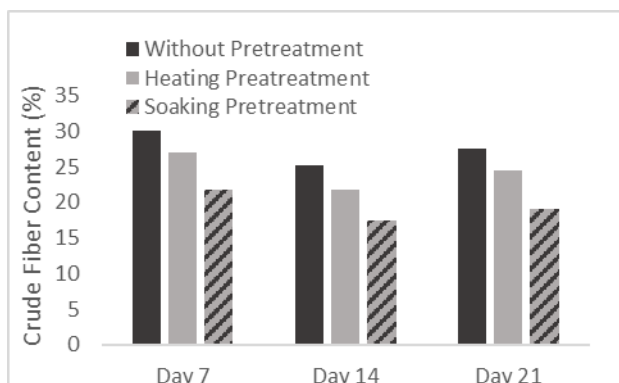


Fig. 2. Correlation of NaOH Pretreatment vs. Crude Fiber Content in Variables with Addition of *Trichoderma* sp.

From both figures it can be seen that the type of soaking pretreatment is more effective to decrease the crude fiber content in solid sago waste followed by heating pretreatment and without pretreatment. Choosing the right pretreatment method is very important because it determines which hydrolysis and fermentation methods will be used later [19]. The uses of strong base solution in this study aims to make the solid sago waste can undergo a delignification process before going into the fermentation process. Delignification of solid sago waste aims to damage the structure of lignin and a part of hemicellulose and also swelling of cellulose structures [20]. Saponification of ester bonds between lignin and hemicellulose residues makes cellulose more open and easier to interact with enzymes, and can decrease the degree of polymerization and crystallinity of cellulose structures [21]. This causes the crude fiber content that has passed the pretreatment stage to be lower than its original level.

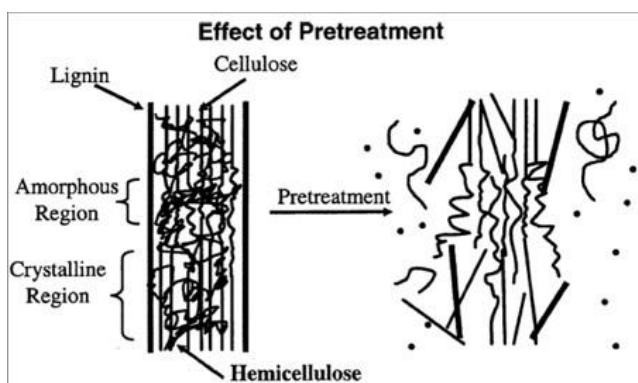


Fig. 3. Schematic process of destruction of lignin structure [22].

Compared with acid pretreatment, the base pretreatment method is more effective as lignin solubilization as presented in Figure 3. Base pretreatment methods in lignocellulose biomass processing are commonly use sodium, potassium, calcium, and ammonium hydroxide. The uses of a base solution cause changes in the structure of lignin by degrading the ester and glycoside side chain. The uses of base solution also cause partial decrystallization of cellulose, partial solubility of hemicellulose and resulting in enlarged cellulose. This process is carried

out by soaking the biomass in an alkaline solution at a predetermined temperature and time. The loosened lignin binds to the Na⁺ ion to form soluble sodium phenolate. Dissolved lignin is characterized by dark color in the solution (black liquor) [23,24]. The neutralization stage needs to be conducted before entering the enzymatic hydrolysis stage to remove lignin and inhibitor substances (eg salts, phenolic acids, and aldehydes) [25]. Pretreatment can increase lignocellulose digestibility in feedstuffs inhibited by several factors, such as: lignin content, cellulose crystallinity, polymerization degree, pore volume, acetyl group bound to hemicellulose, surface area and biomass particle size [26]. The factors that affecting the digestibility of feeds are contents of chemical, protein, fat presentations, fiber and minerals [27].

Figure 4 and 5 shows the correlation of the NaOH pretreatment type and the crude protein content of the feed material derived from the solid sago waste. In Figure 4, the solid sago waste variables without the addition of *Trichoderma* sp on the day 7 samples without pretreatment, heating pretreatment, and soaking pretreatment resulted in crude protein content of 3.96%; 3.12% and 4.05%, respectively, on the day 14 samples without pretreatment, heating pretreatment, and soaking pretreatment yielded crude protein content of 4.04%; 3.18%, and 4.06%, respectively, on the day 21 samples without pretreatment, heating pretreatment, and soaking pretreatment yielded consecutive crude protein content 4.06%; 3.24% and 4.11%, respectively. In Figure 5, the solid sago waste variables with the addition of *Trichoderma* sp on the day 7 samples without pretreatment, heating pretreatment, and soaking pretreatment resulted in crude protein content of 2.80%; 3.38% and 3.54%, respectively, on the day 14 samples without pretreatment, heating pretreatment and soaking pretreatment resulted in crude protein content of 4.06%; 5.99% and 7.96%, respectively, on the day 21 samples without pretreatment, heating pretreatment, and soaking pretreatment resulted in crude protein content of 4.28 %; 6.14% and 8.07%, respectively.

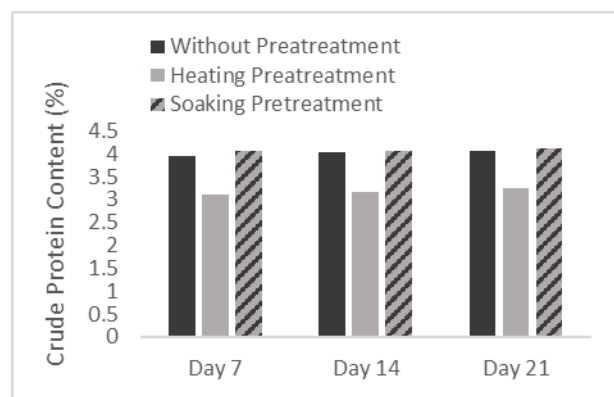


Fig. 4. Correlation of NaOH Pretreatment vs. Crude Protein Content in Variables Without Addition of *Trichoderma* sp.

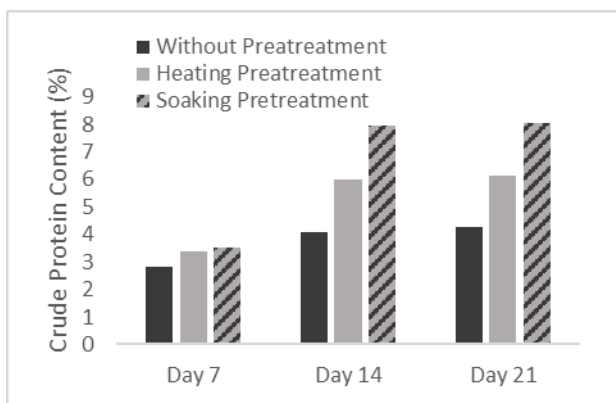


Fig. 5. Correlation of NaOH Pretreatment vs. Crude Protein Content in Variables with Addition of *Trichoderma* sp.

From both figures it can be seen that soaking pretreatment is best way to help raise the crude protein content in the solid sago waste followed by heating pretreatment and without pretreatment. Delignification of solid sago waste aims to damage the structure of lignin and swelling of cellulose structures [20]. According to Saparianti [28], lignin which is a substance of lignocellulose which is a component of crude fiber will prevent enzyme penetration into substrate. In the variable solid sago waste with the addition of *Trichoderma* sp, the presence of delignification can help the work of protease enzymes which are one of the enzymes that produced by *Trichoderma* sp to form protein compounds [29]. Solid sago waste that not added *Trichoderma* sp and without pretreatment procedure tends not to affect the crude protein content. In the variable with the pretreatment of heating at 60°C without *Trichoderma* sp there was a decrease in the crude protein content due to the nature of the protein having denaturation at 50 to 80 °C. Each temperature increase of 10 °C, protein denaturation rate increased to 600 fold [30]. The hydrogen bonds and hydrophobic interactions of non-polar ions of proteins can be damaged by heat.

According to Ophar [30] the kinetic energy that increase due to high temperatures can cause protein molecules to move or vibrate faster, thus breaking the bonds of the molecule. In addition, heat energy will lead to disconnection of non-covalent interactions that exist in the natural structure of the protein but do not break its covalent bond in the form of peptide bonds. Increased the content of crude protein in livestock feed consumed by livestock are good for livestock in terms of nutritional fulfillment. The quality of livestock feed depends on the composition of the nutrients contained therein mainly against dry matter, crude protein, crude fat, crude fiber and feed grade digestibility [31].

3.2 Effect of the addition of *Trichoderma* sp on Crude Fiber and Crude Protein Content

Figure 6, 7, and 8 shows the correlation of the effect of the addition of *Trichoderma* sp to the crude fiber content of the solid sago waste. Figure 6 shows the correlation of the effect of the addition of *Trichoderma* sp vs. crude

fiber content to the solid sago waste variable without pretreatment. Figure 7 shows the correlation of the effect of the addition of *Trichoderma* sp vs. crude fiber content on the variable of solid sago waste with heating pretreatment. Figure 8 shows the correlation of the effect of the addition of *Trichoderma* sp vs. the crude fiber content to the solid sago waste variable with soaking pretreatment. In figure 6, the solid sago waste without pretreatment (without the addition of *Trichoderma* sp and with the addition of *Trichoderma* sp) resulted in crude fiber content: on the day 7 samples: 33.04% and 30.00%, on the day 14 samples: 33.01% and 32.52%, on the day 21 samples: 33.87% and 27.62%. In figure 7, the solid sago waste with heating pretreatment (without the addition of *Trichoderma* sp and with the addition of *Trichoderma* sp) resulted in crude fiber content: on the day 7 samples: 27.50% and 27.01%, on the day 14 samples: 27.49% and 21.71%, on the day 21 samples: 27.98% and 24.53%. In figure 8, the solid sago waste with soaking pretreatment (without the addition of *Trichoderma* sp and with the addition of *Trichoderma* sp) resulted in crude fiber content: on the day 7 samples: 22.03% and 21.79%, on the day 14 samples: 22.62% and 17.36%, on the day 21 samples: 23.00% and 19.11%.

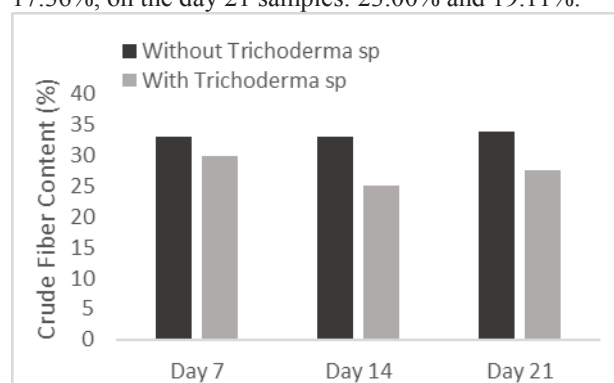


Fig. 6. Correlation of the Addition of *Trichoderma* sp vs Crude Fiber Content to Variables Without Pretreatment.

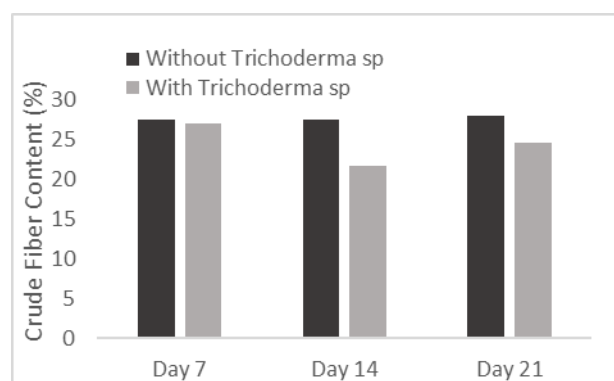


Fig. 7. Correlation of the Addition of *Trichoderma* sp vs Crude Fiber Content to Variables with Heating Pretreatment.

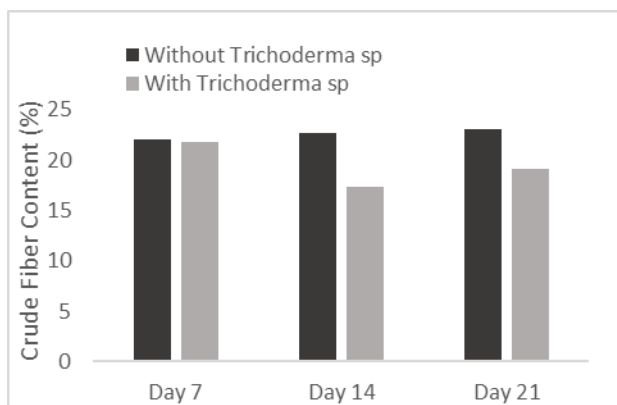


Fig. 8. Correlation of the Addition of *Trichoderma sp* vs Crude Fiber Content to Variables with Soaking Pretreatment.

From the figure 6, 7, and 8, it can be seen that with the addition of *Trichoderma sp* is more effective to reduce the crude fiber content in the solid sago waste compared with without the addition of *Trichoderma sp*. *Trichoderma sp* is a group of complete cellulase-producing soil fungi as well as other components required for total hydrolysis of crystalline cellulose [32]. Cellulase enzyme contains the largest components of cellobiase and β -1,4-glucan-cellobiohydrolase (C1), and β -1,4-glucan-cellobiohydrolase (Cx) in small amounts. This enzyme is hydrolytic and works either consecutively or simultaneously. Cellobiohydrolase is an enzyme that has an affinity for high level cellulose capable of breaking crystalline cellulose while endoglucanase works on amorphous cellulose. Cellobiohydrolase breaks down cellulose by cutting the hydrogen bonds that cause the glucose chains to be readily hydrolyzed. Further hydrolysis takes place to obtain selobiose and eventually glucose by β -glucanase and β -glucosidase enzymes. This series of cellulose hydrolysis reactions causes the crude fiber content of the solid sago waste to be given *Trichoderma sp* to be lower [33]. The factors that affecting the digestibility of feeds are contents of chemical, protein, fat presentations, fiber and minerals [27].

Figure 9, 10, and 11 shows the correlation between the effect of the addition of *Trichoderma sp* and the crude protein content of the solid sago waste. Figure 9 shows the correlation of the effect of the addition of *Trichoderma sp* vs the crude protein content on the solid sago waste variables without pretreatment. Figure 10 shows the correlation of the effect of the addition of *Trichoderma sp* vs the crude protein content on the solid sago waste variables with heating pretreatment. Figure 11 shows the correlation of the effect of the addition of *Trichoderma sp* vs the crude protein content on the solid sago waste variables with soaking pretreatment. In figure 9, the solid sago waste without pretreatment (without the addition of *Trichoderma sp* and with the addition of *Trichoderma sp*) resulted in crude fiber content: on the day 7 samples: 3.9578% and 2.8%, on the day 14 samples: 4.0439% and 4.06%, on the day 21 samples: 4.0558% and 4.28%. In figure 10, the solid sago waste with heating pretreatment (without the addition of *Trichoderma sp* and with the addition of *Trichoderma*

sp) resulted in crude fiber content: on the day 7 samples: 3.12% and 3.38% on the day 14 samples: 3.18% and 5.99%, on the day 21 samples: 3.24% and 6.14%. In figure 11, the solid sago waste with soaking pretreatment (without the addition of *Trichoderma sp* and with the addition of *Trichoderma sp*) resulted in crude fiber content: on the day 7 samples: 4.05% and 3.54% on the day 14 samples: 4.06% and 7.96%, on the day 21 samples: 4.11% and 8.07%.

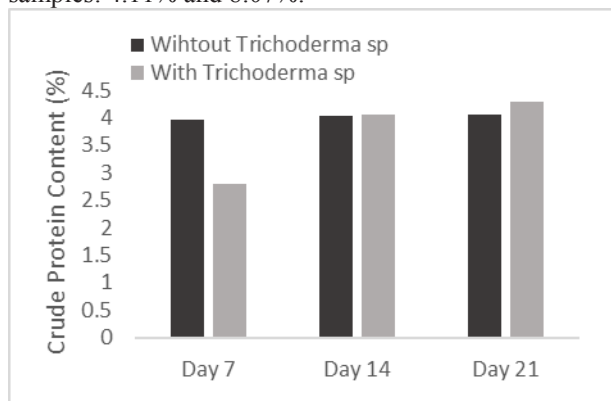


Fig. 9. Correlation of the Addition of *Trichoderma sp* vs Crude Protein Content to Variables Without Pretreatment.

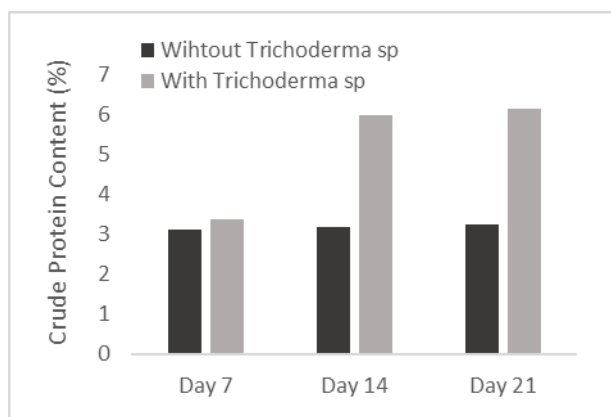


Fig. 10. Correlation of the Addition of *Trichoderma sp* vs Crude Protein Content to Variables with Heating Pretreatment.

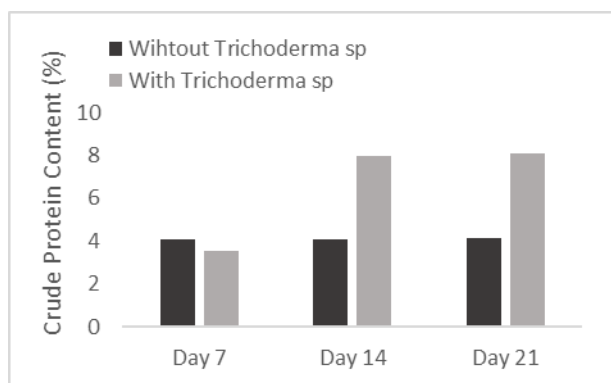


Fig. 11. Correlation of the Addition of *Trichoderma sp* vs Crude Protein Content to Variables with Soaking Pretreatment.

From figure 9, 10, and 11, it can be seen that with the addition of *Trichoderma sp* is more effective to raise the crude protein content in solid sago waste than with no

addition of *Trichoderma* sp. The uses of microorganisms can alter the structure of components and increase enzymatic hydrolysis, added microorganisms can help cellulose hydrolysis process into glucose [34, 35]. *Trichoderma* sp is not only produces cellulose enzymes but also produces protease enzymes that serve to break down proteins [29]. The protein is converted into a polypeptide, and then becomes a simple peptide which ultimately undergoes a further reshuffle into amino acids, which will be exploited by microorganisms to multiply itself. Increasing the number of colonies of microorganisms that are single cell proteins during the fermentation process indirectly increases the crude protein content of the substrate [27, 36]. On the variables without the addition of *Trichoderma* sp there was no significant change in crude protein content.

3.3 Effect of Fermentation Time on Crude Fiber and Crude Protein Content

Figure 12 and 13 shows the correlation of the fermentation time and the crude fiber content of solid sago waste. Figure 12 shows the correlation of fermentation time vs. the crude fiber content of the solid sago waste without the addition of *Trichoderma* sp while figure 13 shows the correlation of fermentation time vs. the crude fiber content of the solid sago waste with the addition of *Trichoderma* sp. From both figures it can be seen that on the day 7, almost all samples except the sample without pretreatment and without the addition of *Trichoderma* sp had decreased contents of crude fiber from the initial level of 33.37%. Then, on the day 14 all samples with the addition of *Trichoderma* sp continued to decrease the crude fiber content while all samples without the addition of *Trichoderma* sp tended not to increase or decrease the crude fiber content. On the day 21, samples with the addition of *Trichoderma* sp have elevated the crude fiber content while samples without the addition *Trichoderma* sp remain constant.

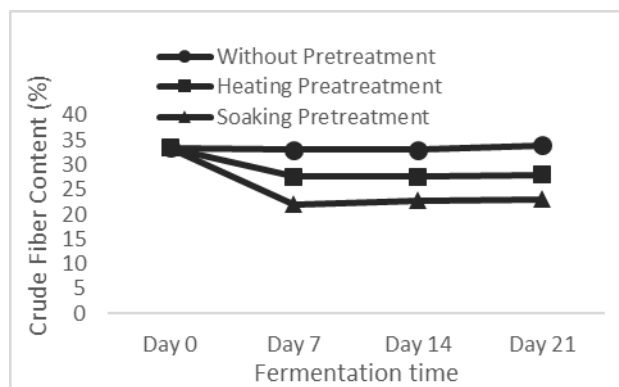


Fig. 12. Correlation of Fermentation Time sp vs Crude Fiber Content to Variables with Addition of *Trichoderma* sp.

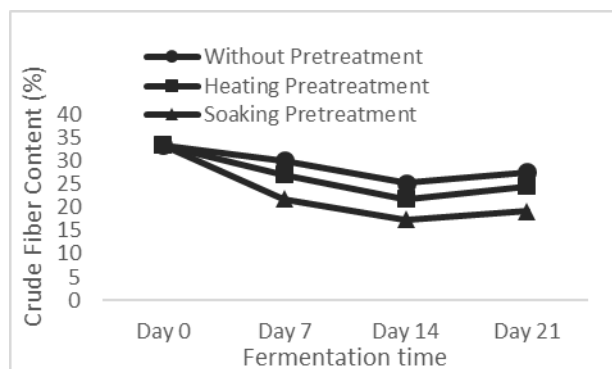


Fig. 13. Correlation of Fermentation Time sp vs Crude Fiber Content to Variables Without Addition of *Trichoderma* sp.

According to Saparianti [28] the longer the fermentation process takes place, then the glucose content produced from the reshuffle of cellulose is higher. This shows the longer the fermentation, cellulose hydrolytic process becomes more effective in lowering crude fiber content. This statement is reinforced by Detroy et al (1981) in [37] which explain that during the first 15 days of fermentation, cellulase-producing microorganisms can increase 2-3 times of baseline reducing sugar. The longer the fermentation means the total mold produced is also higher. Mold will multiply itself in a certain time interval so that the longer the fermentation then *Trichoderma* sp opportunity to multiply itself becomes larger so that the number of cells produced higher. The number of cellulose adsorbed by cellulase depends on the surface area and cellulase concentration. The cellulase adsorbtion on the cellulose surface is usually faster than the overall hydrolysis rate [28]. This causes the hydrolytic reaction to occur more quickly so that the crude fiber content in the solid sago waste becomes lower. This phenomenon occurred from day 0 to day 14 of fermentation on variable solid sago waste with addition of *Trichoderma* sp.

On the day 14 until the day 21 of fermentation there is an increase in the crude fiber content of the solid sago waste with the addition of *Trichoderma* sp. This is due on after the day 14 *Trichoderma* sp has entered the death phase. The cell wall of the dead *Trichoderma* sp is calculated as crude fiber. Meanwhile the solid sago waste variables without the addition of *Trichoderma* sp, the phenomenon that occurs is a decrease in crude fiber content on the day 7 except in the variable without pretreatment. There was no significant change on the day 14 and 21 compared to the previous day. This may result from the apparent delignification of the sample seen significantly on the day 7 which results in a decrease in crude fiber content.

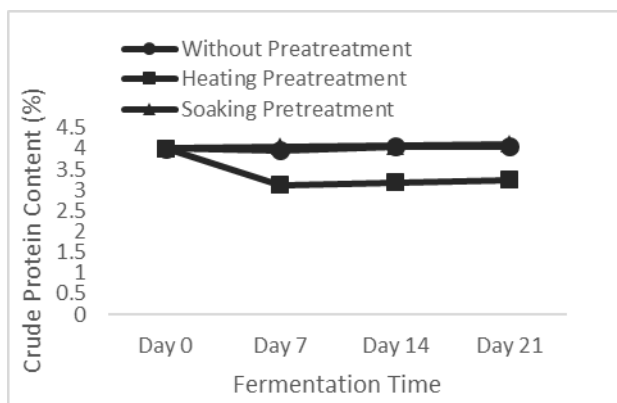


Fig. 14. Correlation of Fermentation Time sp vs Crude Protein Content to Variables with Addition of *Trichoderma* sp.

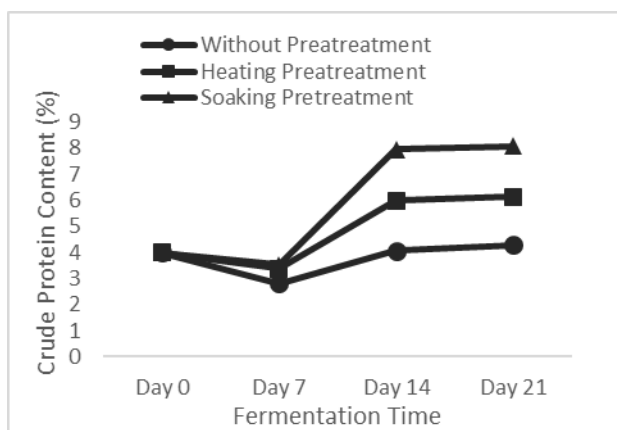


Fig. 15. Correlation of Fermentation Time sp vs Crude Protein Content to Variables Without Addition of *Trichoderma* sp.

Figure 14 and 15 shows the correlation of the fermentation time and the crude protein content of solid sago waste. Figure 14 shows the correlation of fermentation time vs. the crude protein content of the solid sago waste variables without the addition of *Trichoderma* sp, while figure. 15 show the correlation of fermentation time vs. the crude protein content of the solid sago waste variables with the addition of *Trichoderma* sp. From both figures it can be seen that on the day 7, all samples except the sample without pretreatment without addition of *Trichoderma* sp and the sample through pretreatment by soaking without *Trichoderma* sp had decreased crude protein content from baseline. Then, on the day 14 all samples with the addition of *Trichoderma* sp increased significantly in crude protein content while all samples without *Trichoderma* sp did not increase or decrease in crude fiber content. On the day 21, all samples either with the addition of *Trichoderma* sp or without the addition of *Trichoderma* sp did not change the crude protein content.

In the solid sago waste variables with the addition of *Trichoderma* sp on the day 7, the decrease is due to the protein contained in the solid sago waste consumed by *Trichoderma* sp for its growth. Then on the day 14, the phenomenon that occurs is the activity of formation of new proteins from *Trichoderma* sp through protease enzymes. According to Supriyati [17], the best fermentation length is 8-12 days where *Trichoderma* sp

increases the crude protein content of the substrate. The protein is converted into a polypeptide, and then becomes a simple peptide which ultimately undergoes a further reshuffle into amino acids, which will be exploited by microorganisms to multiply itself. Increasing the number of colonies of microorganisms that are single cell proteins during the fermentation process indirectly increases the crude protein content of the substrate [27, 36].

The difference in levels of the crude protein content in sample added with *Trichoderma* sp was suspected because the crude fiber content of the solid sago waste had been reduced previously through the delignification process. Delignification of sago pulp aims to damage the lignin structure and swell the cellulose structure [20]. According to Saparanti [28], lignin which is a substance of lignocellulose which is a component of crude fiber will prevent enzyme penetration into substrate. The decrease in crude fiber indicates that the enzyme penetration process into the substrate is better and capable of producing higher the crude protein content.

The life phases of microorganisms in the substrate affect the content of crude fiber and crude protein [38]. The phase is generally influenced by the type of microorganisms used and the type of substrate used. The life phase of the organism is divided into 5: adaptation phase, initial growth phase, fast growth phase (logarithmic), stationary phase, and death phase. Adaptation phase is the initial period and is a phase of adjustment (adaptation), so there is no increase in the number of cells and sometimes the number of cells decreases. In the early growth phase there was no population increase. Cells undergo changes in chemical composition and increase in size and the intracellular substance increases. Logarithmic phase is a rapid multiply period. In this period can be observed features of active cells. The generation time on each microorganism can be determined in this rapid phase. At that phase it can be seen that some cells begin to divide, others half divide, and others finish splitting. In the static phase the breeding begins to decrease and some cells die. If the rate of multiply is equal to the rate of death, then the total number of cells remains constant. This may be due to reduced nutrients or the formation of metabolic products that tend to accumulate may be toxic to microorganisms. The death phase is the phase in which the multiply process has stopped. The cells are dead, which will then be followed by a lysis process. If the rate of death exceeds the rate of multiply, then the actual number of cells decreases [38].

4 Conclusions

Livestock feed production using solid sago waste processed through soaking pretreatment and fermentation with *Trichoderma* sp yields data of crude fiber content decrease from 33,3734% to 17,363% and increase of crude protein content from 4% to 7,955% at 14 days of fermentation time. This indicates the

optimum enzyme productivity in the growth phase of *Trichoderma* sp in 14 days.

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