

Fig 2. Structure of C-10(6-prenyloxycoumarins)

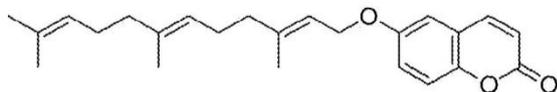


Fig 3. Structure of C-15(6-prenyloxycoumarins)

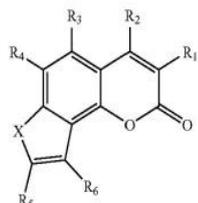


Fig 4. Structure of coumarin/chalcone

Since coumarin showed interesting antiproliferative or anticancer activity, we decided to investigate *Calophyllum* species led to the isolation of one chemically distinct group of coumarin. In these studies we are describing the antiproliferative activity of the phenylcoumarin to MCF-7 cell lines from *C. incrasaptum*.

2 Materials and methods

2.1 Materials

Stem bark of *C. incrasaptum* M.R Henderson- Wyatt Smith was collected from Kalimantan Island in Indonesia. The sample was identified at the Herbarium Unit, Department of Botany Indonesian Institute of Sciences (LIPI). Solvents and chemicals used in this study are analytical grade and were purchased from Merck (Darmstadt, Germany).

2.2 General aspects

¹H and ¹³C NMR spectra were recorded on a JEOL NMR spectrometer at 500 and 125 MHz, respectively. The abbreviations s, d, and br used to describe ¹H spectra refer to singlet, doublet, and broad, respectively. NMR spectra were recorded in CDCl₃ solution on a Jeol JNM ECA 500 MHz instrument, using TMS as the internal standard. Silica gel (60–230 mesh and 230–400 mesh) were used for column chromatography and preparative TLC plates coated with silica gel (2000 micron thickness) was purchased from Merck (Darmstadt, Germany). Precoated Si gel plates SIL G/UV₂₅₄, 0.25 mm were used for TLC. The compounds were detected under UV light at 254 and 366 nm. Melting points were determined on an Electrothermal Fisher melting point apparatus (Scientific serial 903N0056) and are uncorrected. IR spectra were

recorded on a FTIR Prestige 21 Shimadzu spectrophotometer (KBr dish) and UV spectra was taken on a Hitachi U-2000 spectrophotometer. LC-MS were recorded on Mariner Bio Spectrometric equipment with pneumatically assisted electrospray ionization (EIS) source.

3 Methods

3.1 Extraction procedure

The air-dried stems bark of *C. incrasaptum* (2.1 kg) were minced finely and shredded in an electric mill, macerated three times with MeOH (6L x72h weight of extract after solvent was removed : 400 gr), then was diluted in H₂O and partitioned with between *n*-hexane and H₂O: EtOAc and H₂O : and butanol – H₂O . The *n*-hexane, EtOAc and butanol was removed under reduced pressure to give 65.8 g, 138.8 g and 609.4 g *n*-hexane, EtOAc and BuOH extract respectively.

EtOAc extract (120 gr) was separated by flash column chromatography, It was eluted with a stepwise gradient of *n*-hexane followed by increasing amounts of EtOAc (20:1; 10:1; 5:1; 3:1; 1:1 ; EtOAc 100%) give 6 fractions (Fr. A1–A6), then were eluted by EtOAc:MEOH (20:1; 10:1; 5:1; 3:1 ,1:1) to give 5 fractions (Fr A7-A11). Fraction A5 was further purified several times by normal CC system on silica gel medium-pressure liquid chromatography (MPLC), eluting with *n*-hexane and 10% stepwise gradient of acetone to afford several fractions, fractions were collected and regrouped on the basis of TLC analysis. Sub fraction 68-74 was then applied to a silica column eluted with *n*-hexane – DCM (dichloromethane 50:50→0:100). Fraction 3 was further purified by sephadex LH-20 , approximately 15 ml was collected for each fraction, collection sub fraction 33 - 47 were pooled after TLC profile analysis. We were able to obtain a pure crystal of the phenyl coumarin by recrystallizing method using *n*-hexane : acetone (3:1).

3.2 Preparation of Cancer Cells Line

Cancer cells line used in this study is breast carcinoma (MFC-7), and the cell line was cultured in DME Medium with FBS 10%. These cells were cultured at 37°C with moisture content of 95% and 5% CO₂ for 3 days until the cell cultures undergo confluence 60-70%. After that the old media removed, replaced with new medium and incubated again for 24 hours. Culture cells then washed with PBS 1-2 times as much and was suspended using trypsin-EDTA solution. Cells that have been suspended added with new.

3.3 In vitro antiproliferative assay

Antiproliferative assay was used by Alamar Blue method in triplicate against human breast carcinoma (MFC-7) cell lines. The MCF culture cells line of 100 μL was supplemented with 10 μL of pure test compound (05; 1 and 1.5 ppm). It was then incubated for 24 hours at a 37°C.

Colouring process was done by adding Alamar blue solution for 4 hours. The colour intensity of the cell line was measured by using ELISA plate reader at 560 nm (excitation) and 590 nm (emission) wavelength. Percent viability is calculated as in equation 1. While IC₅₀ of the active extract was calculated by linear regression analysis between percent survival showed in equation 2. Alamar Blue was used to color the sample, and the IC₅₀ of the pure compound (Alamar Blue Assay, U.S. Patent No.5,501,959). [12]

$$\% \text{ Viability} = \frac{OD(\text{cell+sample}) - OD(\text{negative control})}{OD(\text{cells}) - OD(\text{negative control})} \times 100$$

4 Result and Discussion

4.1 Results of Elucidation structure

Phenylcoumarin was isolated via column chromatography (CC) as an yellow amorphous from an EtOAc extract of steam bark of *C. incrasaptum* M.R Henderson- Wytt Smith, mp 184-185°C. The UV absorbtion of phenylcoumarin characterized a 5,7,10 threeoxygenated coumarin, and give a positive test with a vaniline/H₂SO₄ reagent, an intense deep blue spot develops over 24 hour period. LC-MS revealed the molecular formula to be C₂₁H₁₈O₅ (*m/z* 351,40 [M+1]⁺), by ESIMS analysis, i.e., the molecule had threeteen degrees of unsaturation. The UV spectrum exhibited two maxima at λ_{max} 256 and 319 nm were indicative of a dimethylchromanone derivative, whilst the FTIR spectra showed important peaks explaining the bending, stretching, and double-bond absorptions of the coumarin. The C-H stretching of absorption for CH₃ occurred at wavelengths of 2937 cm⁻¹. One alkane peaks were observed showing the bending absorption for CH₃ (ν_{max} 1332 cm⁻¹). spectrum indicated absorption bands ν_{max} 1732 cm⁻¹ attributed to non-conjugated carbonyl group (C=O), and ν_{max} 1691 cm⁻¹ as conjugated C=O carbonyl groups, ν_{max} 1597-1332 (aromatic C=C) and ν_{max} 1153 cm⁻¹ (C-O), and mono substituted benzene ring at ν_{max} 767 and 702 cm⁻¹ (Figure 5).

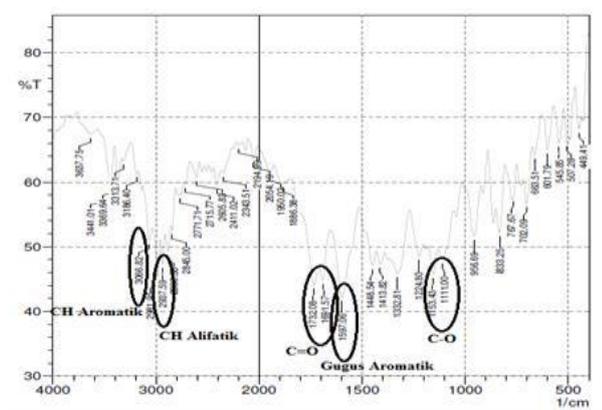


Fig 5. IR spectrum of isolate compound

Determination of structure was performed using a NMR device. The 1-NMR spectrum of isolate compound

(phenyl coumarin) is evidenced by the presence of two proton singlet (δ_H 6.07, 1H, s, H-3) and δ_H 6.73, 1H, s, H-6) and group of signal belonging to five aromatic proton of the phenyl group [(δ_H 7.34-7.40, *m* : δ_H = 7,34 (1H, *m*), 2 protons at δ_H 7.40 (2H, *m*), and 2 protons at δ_H 7.38 (2H, *m*)]. The presence of a singlet-shaped signal at δ_H 3.14 ppm (3H, s, H-13) shows the proton of one methoxy group. Addition signal included these of dimethylchromanone ring (δ_H 4.28, 1H, *m*, H-8, and δ_H 2.55, 1H, *m*, H-9 : δ_H 1.52, 3H, *d*, *J* = 6.5 Hz, H-11 and δ_H 1.18, 3H, *d*, *J* = 7.1 Hz, H-12) (Figure 6).

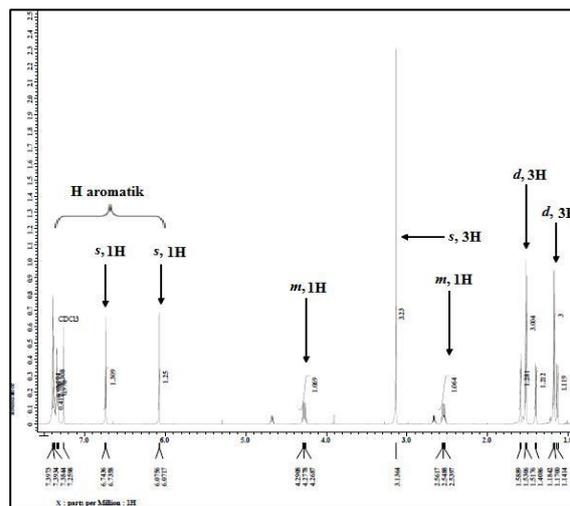


Fig 6. ¹H-NMR spectrum of isolate compound

The ¹³C-NMR data shows 21 peaks of carbon atoms, nine C quaternary atoms (including two carbonyl downfield signals at δ 159.43 and 191.65), seven C-H aromatic, two aliphatic protons (δ_H 1.52, *m*, H-8 & δ_H 2.55, *m*, H-9), the presence of two methyl groups (CH₃) is supported by ¹³C-NMR data with a carbon chemical shift of 19.65 and 10.49 ppm and one methoxy group at 62.77 ppm (Figure 7).

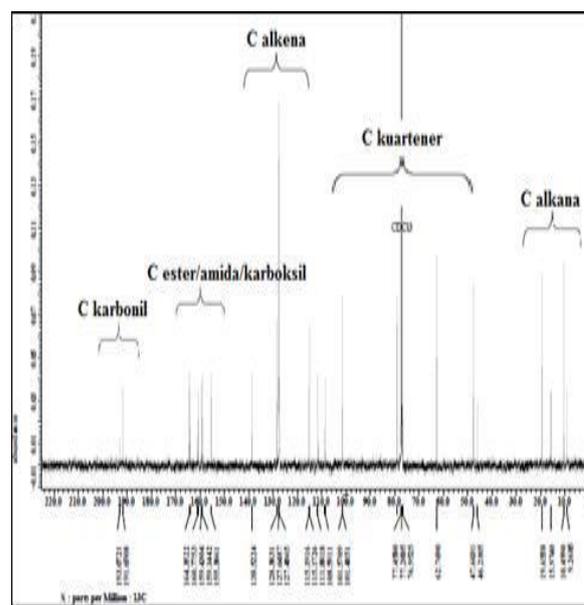


Fig 7. ¹³C-NMR spectrum of isolate compound

This phenyl coumarin compound have a methoxy substituted at C-5 (singlet methyl signal at δ_H 3.14 ppm). The NMR signals observed were in accordance with the presence of coumarin nucleus, with the benzenoid ring (δ_C 159.43, 115.17, 155.38, 108.59, 160.77, 101.48, 164.35, 111.88 and 159.14) an unsubstituted phenyl ring (138.12, 128.38, 127.49 and 127.65) one a methoxyl group ($\delta_C = 62.77$) and a coumarone ring ($\delta_C = 191.5, 78.99, 47.68, 19.65$ and 10.49). A correlation from the phenyl C1' (δ 138.2) to H-3 confirmed that a phenyl group was attached to C-4 as in isolated compound.

Long-distance 3J and 2J (HMBC) proton H 6,07 ppm on C-3 has been proved by H-correlation at δ_H 6.07 with C-4a (δ_C 108,59), C-2 (δ_C 155,43) and C-1' (δ_C 138,52), this proves the position of H 6.07 ppm at C-3. The presence of two doublet-shaped signals at δ_H 1.52 ppm (3H, *d J*, 6.5 Hz) and 1.18 ppm (3H, *d J*, 7.1 Hz) show protons of two methyl groups. HMBC data for proton correlation of methoxy group ($\delta_H = 3.14$ ppm) with carbon C-5 (δ_C 160,77) assured the position of methoxy on C-5.

Phenyl group in C-4 positions is evidenced by the correlation of proton aromatics δ_H 7.38 ppm with carbon at δ_C 155.38 ppm, and proton correlation δ_H 6.07 ppm (H3) with carbon at δ_C 138.52 ppm (Figure 8). HMBC correlation of isolate compound was explained in figure 8.

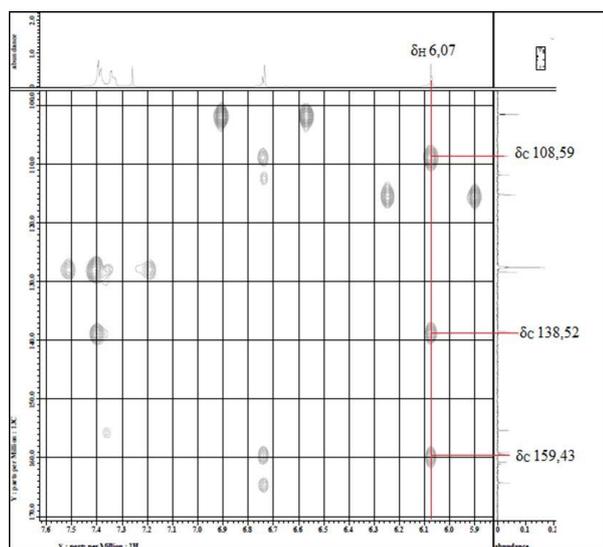


Fig 8. HMBC spectrum of isolate compound

All NMR 1D and 2D data was reported in Figure 9 and Table 1.

Table 1. Data NMR 1D and 2D and COSY chemical shift (ppm) of phenyl coumarin

No	C	H-NMR (δ_H) ppm	Cosy
1	0		
2	159.43	-	
3	115.17	6.07 (1H,s)	-
4	155.38	-	-
4a	108.59	-	-

5	160.77	-	-
6	101.48	6.73 (1H,s)	-
6a	164.35	-	-
7	0	-	-
8	78.99	4.28 (1H, <i>m</i> , <i>J</i> =6.5 Hz)	H-8 - H9
9	47.68	2.55 (1H, <i>m</i> , <i>J</i> =6,5 Hz)	H-9- H-8
10	191.65	-	-
10a	111.88	-	
10b	159.14	-	-
11	19.65	1.52 (3H, <i>d</i> , <i>J</i> =6.5 Hz)	H11-H8
12	10.49	1.18 (3H, <i>d</i> , <i>J</i> = 7.1 Hz)	H12-H-9
1'	138.2	-	-
2'	128.38	7.38 (1H, <i>m</i>)	H-2'-H-3'
3'	127.65	7.40 (1H, <i>m</i>)	H-2'-H-4'
4'	127.49	7,34 (1H, <i>m</i>)	H-3'-H5'
5'	127.65	7.40 (1H, <i>m</i>)	H-4'. -6'
6'	128.38	7.38 (1H, <i>m</i>)	H-6'-H-5'
OCH ₃	62.77	3.14 (3H, s)	

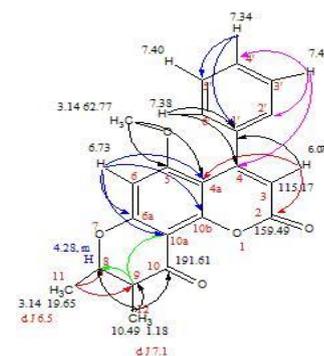


Fig 9. HMBC correlation of of isolate compound

This paper report the crystal structure of one isolated compound from *C. incrasaptum* as phenyl coumarin (Figure 10).

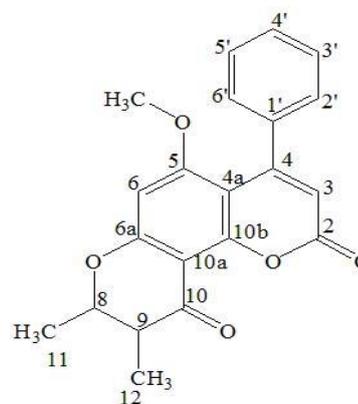


Fig 10. Chemical structure of phenyl coumarin

Hormon oestrogen has the crucial role in development of the breast cancer, the most frequent malignant disease in women, therefore many therapies are designed to block

this activity. Cinnamoyl-coumarin derivatives were especially effective in estrogen-dependent cancer, such as breast (MCF7) cancer cell line. The phenylcoumarin in these paper have anticancer activity too, because coumarin are proven to posses a wide range of biological activitied , e. i as anticancer.

4.2 Results of cell viability

Inhibition of cell viability caused by prenyl coumarins including phenylcoumarin from *C. incrasaptum* was examined using Alamar Blue assay. The cytotoxic effect of pure test compound (phenyl coumarin) against MCF-7 breast cancer cells was determined with alamar blue assay at range of 0-1.5 ppm after 24 h of treatment period. The cell viability can be qualitatively assessed by the color intensity of alamar blue. The intensity of the alamar blue dye increased when the concentration of pure test compound decreased. The results are graphically represented in Figure 11. Cell viability (%) was observed to be conversely proportioned to the concentration of the pure test compound, these results showed that these phenylcoumarin compound indicated decreased cell viability (that treatments showed growth inhibition) cel MCF7 responded in a dose-dependent. The pure test compound showed effective cytotoxic effect with the IC₅₀ value of 2,6 ppm in MCF-7 breast cancer cells

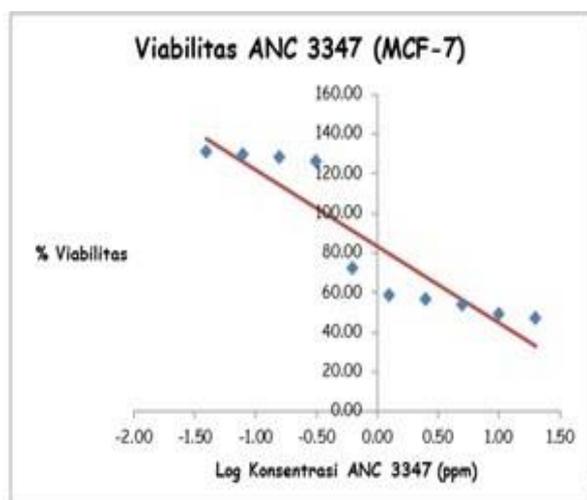


Fig 11. Cytotoxic effects of phenylcoumarin on MCF7 cells. Relationship between log concentration with % viabilirty

lines obtained from regression : $Y = -54.04 \text{ Log } X + 68.80$ with $R = 0.90$ $IC_{50} = 2,23 \mu\text{g/mL}$
 IC_{50} values in ($\mu\text{g/mL}$), $IC_{50} < 4 \mu\text{g/mL}$ is highest cytotoxic,
 IC_{50} 4-30 $\mu\text{g/mL}$ considered cytotoxic and in active,
 $IC_{50} > 40 \mu\text{g/mL}$

4.3 Discussion

According to the results, we treated the MCF7 cell line with different concentrations of phenylcoumarin for 24 h. The results indicated phenylcoumarin induced cell death in concentration –dependent manner.

Phenyl coumarin is a group of compounds which have shown to psses preventive and therapeutic effects on breast cancer.

5 Conclusions

Using a combination of NMR techniques (¹H and ¹³C NMR, HMQC, HMBC and COSY) and IR spectroscopy combined with DEPT calculations, it has been possible to determine the structure of phenyl coumarin (Figure 10)

Phenylcoumarin exerts antiproliferative effect in breast carcinoma cell line, and can be considered for further mechanistic evaluation in human its biosafety and anticancer effect.

The results demonstrated that treatmen of MCF7 cells with phenylcoumarin results in a significant decrease in cell viability, IC₅₀ values against MCF7 is 2.23 $\mu\text{g/mL}$.

The authors would like to thank the LIPI (Indonesian Institute of Sciences) for financial support through Kompetitive project thanks are due to Mr. Ismail Rahman at the Herbarium staff in Research Centre for Biology- LIPI Cibinong for identifying the plant material.

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