

## Bioesterified polysubstituted-cyclodextrin/surfactant nanoparticles obtained by multilevel self-assembly

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**Abstract.** The purpose of this work is to investigate the inclusion complexation between a novel amphiphilic biotransesterified cyclodextrin (CD), incorporated in nanostructured environment, and a model drug compound. A water-insoluble  $\gamma$ -cyclodextrin derivative ( $\gamma$ -CD-C<sub>10</sub>), polysubstituted with multiple ( $n=7-8$ ) decanoyl chains (C<sub>10</sub>) on the secondary face, is produced by enzymatically-assisted esterification. The  $\gamma$ -CD-C<sub>10</sub> derivative is embedded in amphiphilic nanoenvironment created by self-assembly with the lipophilic dye Nile red (NR) and the non-ionic surfactant polysorbate 80 (P80). The inclusion complexation and the environmental effects upon the  $\gamma$ -CD-C<sub>10</sub>/NR/P80 nanoparticle (NP) formation, in a multilevel self-assembly approach, are investigated by means of steady-state fluorescence and Förster resonance energy transfer (FRET) techniques. Quasi-elastic light scattering (QELS) is used to control the NP size distribution during the sequential steps of the assembling process.

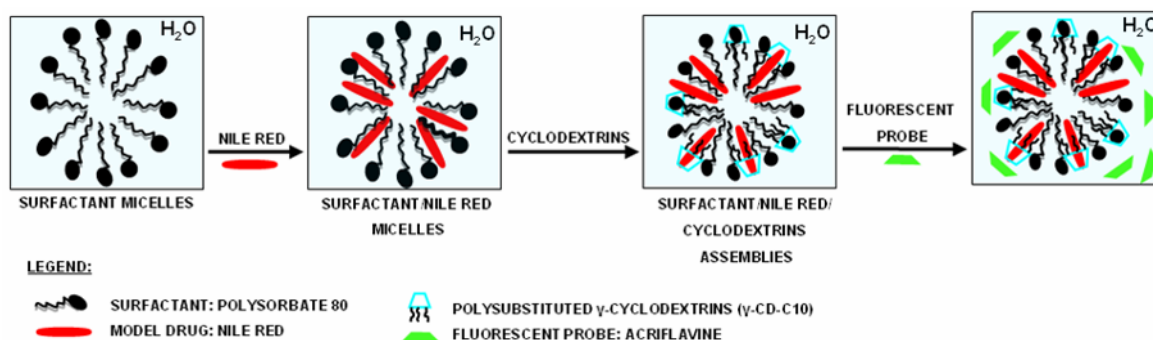
### 1 Introduction

Cyclodextrins (CDs) are recognized host materials capable of inclusion complex formation with guest water-insoluble molecules, which are set in their hydrophobic nanocavities. This feature is being used to improve drug stability and bioavailability by increasing the drug permeability properties, solubility and/or dissolution through inclusion complexation, as well as to reduce the drug toxicity and side effects [1-9]. The formation of complexes between CDs and drug molecules *via* dynamic noncovalent interactions, in a host-guest model, has drawn considerable interest [1,9-12].

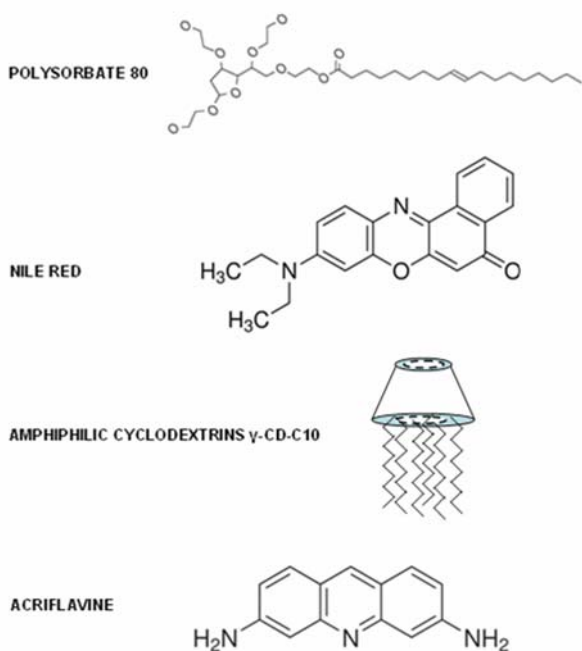
A recent methodology for grafting of alkyl chains on the secondary face of native CDs by enzymatic transesterification, using thermolysin as a biocatalyzer, has opened new possibilities for the design of drug nanocarriers with colloidal behaviour [8,9]. Amphiphilic CD derivatives with grafted decanoyl chains (CD-C<sub>10</sub>) may self-assemble into nanospheres with multilamellar

inner organization or into reservoir-type particles, which can embed lipophilic drugs [5,7-9]. However, the unmodified CD-C<sub>10</sub> particles may readily aggregate depending on the surroundings. At variance, the modification of the CD-C<sub>10</sub> particles by nonionic surfactants, like polysorbate 80 (P80) (which involves poly(ethylene glycol) moieties in the polar head-group region) is anticipated to increase their steric stability, bioavailability, and cellular uptake. Towards this possibility, we here create mixed amphiphilic  $\gamma$ -CD-C<sub>10</sub>/P80 assemblies and study the inclusion complexation mechanism with a fluorophore substance employed as a model drug.

The steps of the multilevel assembly approach leading to formation of polysubstituted-CDs/surfactant/model-drug nanoparticles (NPs) are schematically presented in Fig. 1. The chemical structures of the investigated compounds are shown in Fig. 2. Nile red (NR) was chosen as a model drug (interacting with the CD cavity) because of its hydrophobic fluorophore and



**Fig.1.** Scheme of the multilevel self-assembly process at a low molar content of  $\gamma$ -CD-C<sub>10</sub>.



**Fig.2.** Structural formulae of the investigated molecules.

its stable fluorescence. NR has been previously studied as a polarity-sensitive fluorescence probe suitable for monitoring of structural transformations and intermolecular interactions upon micropolarity changes in the surroundings [13,14].

The formation of an inclusion complex between the  $\gamma$ -CD-C<sub>10</sub> molecules and the guest fluorophore NR is studied here by fluorescence techniques. Fluorescence and UV-Visible spectroscopies have been appropriate methods for establishing the formation of inclusion complexes between CDs and fluorophores [15-17]. Here nanostructured environment of PEGylated micelles was prepared using the non-ionic surfactant P80 (Fig. 2). NR was added to the medium in order to form micelles incorporating the fluorophore (Fig. 1). Then, different amounts of cyclodextrins ( $\gamma$ -CD-C<sub>10</sub>) were added in order to study the complex formation between the lipophilic NR and the CD cavities in nanostructured surroundings. Förster resonance energy transfer (FRET) was investigated between the donor dye acriflavine (AC) (Fig. 2), included in the aqueous phase, and the acceptor NR dye incorporated in the surfactant micelles or in the  $\gamma$ -CD-C<sub>10</sub>-containing nanoassemblies.

## 2 Experimental section

### 2.1 Materials

Polysorbate 80 (P80), Nile red (NR), and acriflavine (AC) were purchased from Sigma-Aldrich. The amphiphilic  $\gamma$ -cyclodextrin derivative,  $\gamma$ -CD-C<sub>10</sub>, was enzymatically synthesized as described by Choïnard et al. [9]. The average molecular weight,  $M_n$ , of the  $\gamma$ -CD-C<sub>10</sub> ester was 2505 g/mol as determined by MALDI-TOF analysis.

### 2.2 Micelles preparation

100 ml of P80 micellar solution was prepared at concentration 10 mM in pH 7.4 phosphate buffer (solution A). 0.036 g of hydrophobic NR was added to 50 ml of solution A. This yielded 2.25 mM dye concentration in P80 micellar medium (solution B).

### 2.3 Preparation of $\gamma$ -CD-C<sub>10</sub>/P80 mixed assemblies incorporating the guest NR

NPs dispersions were prepared at different molar ratios of  $\gamma$ -CD-C<sub>10</sub> with regard to P80. Briefly, 2.5 mg, 5 mg, 7.5 mg or 10 mg of  $\gamma$ -CD-C<sub>10</sub>, corresponding to 0.9 mol%, 1.8 mol%, 2.9 mol%, and 3.5 mol% of cyclodextrins, were added to 14900  $\mu$ l of solution A (solution C). Then, 100  $\mu$ l of solution B were added to solution C in order to obtain 15 ml of  $\gamma$ -CD-C<sub>10</sub>/P80 mixed assemblies incorporating the fluorophore NR. The resulting concentration of NR was equal to  $1.5 \times 10^{-5}$  M in this 15 ml preparation. Vortexing and sonication was applied to ensure mixing of the components in all cases.

### 2.4 Fluorescence spectra measurements

Excitation and emission spectra of the two dyes were recorded at room temperature by means of a spectrofluorimeter SPEX Fluorolog (model 1681 0.22 m) with an excitation wavelength of  $\lambda_{ex} = 455$  nm for acriflavine (AC) and of  $\lambda_{ex} = 543$  nm for Nile red (NR). The excitation and emission spectra were normalized in order to display the overlapping of bands between the fluorescence emission of AC and the absorption spectrum of NR. The excitation and the emission spectra of AC were obtained in phosphate buffer (pH 7.4), whereas those of NR were acquired in P80 micelles. The AC and NR concentrations were  $3.3 \times 10^{-8}$  M and  $5 \times 10^{-6}$  M, respectively.

### 2.5 FRET measurements

Steady-state fluorescence spectra measurements were performed with the  $\gamma$ -CD-C<sub>10</sub>/NR/P80 nanoassemblies in the presence of AC (donor dye) in the aqueous phase using the same equipment. Briefly, 1 ml solution of NR-labelled  $\gamma$ -CD-C<sub>10</sub>/P80 mixed micelles, incorporating amphiphilic CDs with a varying molar fraction, was dropped down in a spectroscopy cell containing 1980  $\mu$ l of phosphate buffer (pH 7.4). For the FRET study, 20  $\mu$ l of acriflavine stock solution ( $C_{stock} = 0.05$  mM) were added to the spectroscopy cell. The resulting AC and NR concentrations, in the spectroscopy cell, were of  $3.3 \cdot 10^{-8}$  M and  $5 \cdot 10^{-6}$  M, respectively. FRET was recorded for the AC/NR donor/acceptor pair at room temperature. The excitation wavelength was set at 480 nm and the emission spectra were collected from 500 to 800 nm.

### 2.6 Size measurements

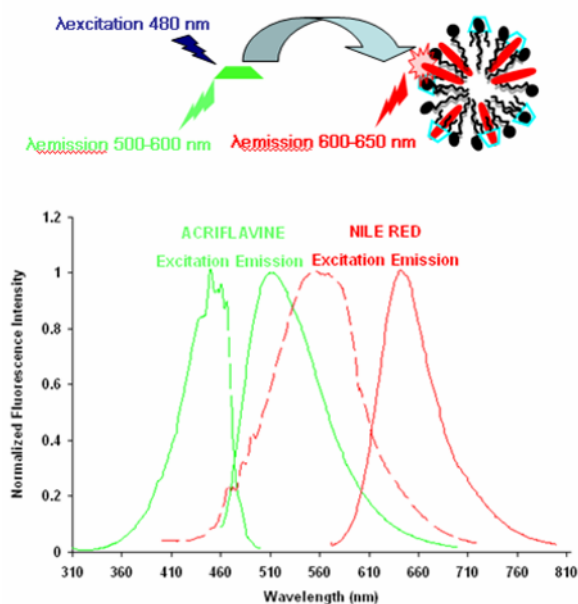
The samples studied by fluorescence spectroscopy were analysed also by quasi-elastic light scattering (QELS). The Zetasizer Nano ZS-90 system was a MALVERN instrument (light source He-Ne laser 633 nm, maximum

power 5 mW). Every sample was measured three times. The scattering angle was fixed at 90° and the temperature was 25°C. The concentrations of the samples were the same as for the fluorescence experiments for either the initial P80/NR micelles or the  $\gamma$ -CD- $C_{10}$ /NR/P80 mixed nanoassemblies.

### 3 Results and discussion

#### 3.1 Fluorescence spectra of the studied dyes

Figure 3 characterizes the AC/NR FRET pair. It shows that the emission spectrum of the donor dye, acriflavine (AC), overlaps with the excitation spectrum of the acceptor dye, Nile red (NR). The established good spectral overlap implies that it should be possible to observe a FRET event from acriflavine to Nile red when the two fluorophores come in a sufficient proximity in the investigated nanostructured system. In fact, FRET occurs when the electronic excitation energy of a donor chromophore is transferred to an acceptor molecule, located nearby, through dipole-dipole interaction occurring in the donor-acceptor pair.



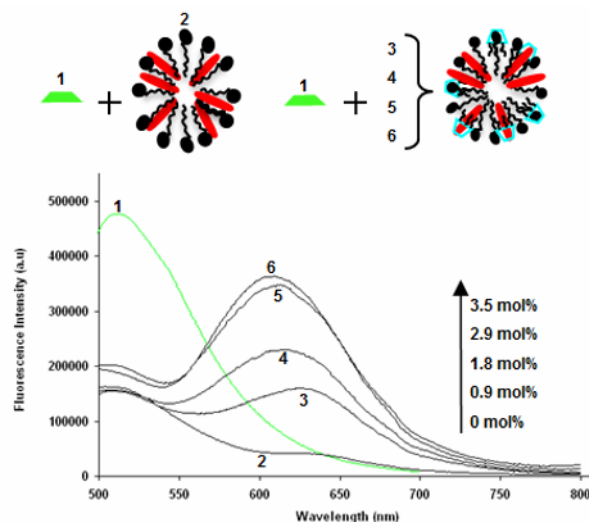
**Fig.3.** Overlapping between the fluorescence emission of the donor dye (acriflavine) in a phosphate buffer and the absorption spectrum of the acceptor molecule (Nile red) in P80 micelles. The excitation wavelengths were 455 nm and 543 nm for acriflavine and Nile red, respectively. The emission spectra were recorded in the range 460-700 nm and 570-800 nm for the two dyes, respectively.

#### 3.2 Mechanism of inclusion complexation in NPs

In the investigated here colloidal nanoassemblies, the components of the FRET pair are located in different environments. AC is water soluble, while NR is incorporated in the mixed assemblies and may be anchored in the nanocavities of the  $\gamma$ -CD- $C_{10}$ . The donor dye AC surrounds the P80/NR micelles as well as the  $\gamma$ -CD- $C_{10}$ /NR/P80 NPs (Fig. 1, right). A greater proximity

of the two dyes, and hence more efficient FRET, would be achieved if the acceptor (NR) is located at the polar/apolar interfaces of the  $\gamma$ -CD- $C_{10}$ /NR/P80 nano assemblies rather than solubilized deeply in the core of the micelles or the NPs formed at increasing CDs content.

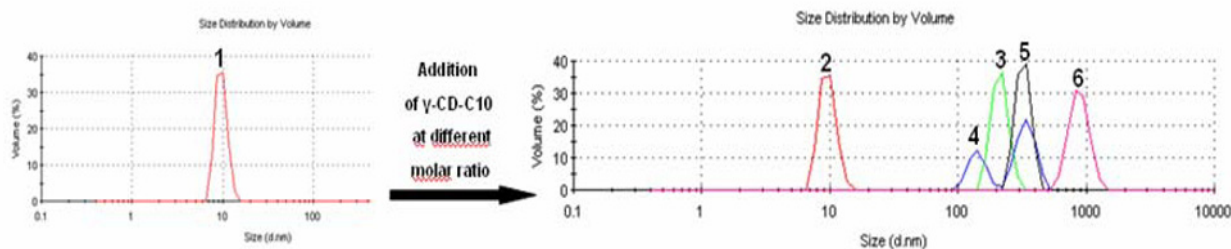
Figure 4 shows that the donor dye AC is strongly fluorescent with a maximum emission at 510 nm. The fluorescence intensity of AC decreases sharply upon the addition of NR-labelled micelles (spectrum 2). The NR fluorescence emission was observed at 630 nm.



**Fig.4.** Fluorescence emission spectra of (1) acriflavine in phosphate buffer (pH 7.4) and in (2) Nile red/P80 micelles. The AC concentration is  $3.3 \cdot 10^{-8}$  M and the NR concentration is  $5 \cdot 10^{-6}$  M. The fluorescence emission maximum of NR is enhanced at increasing concentrations of amphiphilic CDs. The  $\gamma$ -CD- $C_{10}$  molar fraction is varied as (2) 0 mol%, (3) 0.9 mol%, (4) 1.8 mol%, (5) 2.9 mol%, (6) 3.5 mol% with respect to P80 (see Inset). The excitation wavelength for AC is 480 nm.

Spectra 3 to 6 show the progressive increase in the emission maximum intensity of NR upon augmenting the molar fraction of  $\gamma$ -CD- $C_{10}$  molecules in the NPs. The concentration of NR is constant in all cases. Hence, if NR has remained located at the same distance in the mixed structures (3–6) from the AC acceptor, the spectral intensities would be always equal to that in spectrum 2. However, one observes an increase in the fluorescence emission of NR and a blue shift of its fluorescence peak maximum at increasing molar fractions of amphiphilic  $\gamma$ -CD- $C_{10}$  molecules, which present cavities for inclusion complexation. The fluorescence of NR is essentially stronger in the obtained NP assemblies with  $\gamma$ -CD- $C_{10}$  as compared to that in P80 micelles. Evidently, inclusion complexes are formed between  $\gamma$ -CD- $C_{10}$  and NR in nanoconfined geometries.

The increase in the fluorescence emission of NR and the blue shift of the NR peak maximum are more significant at higher molar fractions of  $\gamma$ -CD- $C_{10}$ . This indicates an increased hydrophobicity of the NR surroundings and changes in the micropolarity around the NR fluorophore in the presence of CDs. Altogether, these results confirm that there is an inclusion complex formation between the hydrophobic nanocavities of the CDs and the guest model drug NR.



**Fig.5.** Particle size distributions of polysorbate 80/Nile red micelles (1, red curve) and of polysorbate 80/Nile red/ $\gamma$ -CD-C<sub>10</sub> mixed assemblies with 0 mol% CD (2, red curve), 0.9 mol% CD (3, green curve), 1.8 mol% CD (4, blue curve), 2.9 mol% CD (5, black curve) and 3.5 mol% CD (6, pink curve).

At a low molar fraction of  $\gamma$ -CD-C<sub>10</sub> (< 1 mol%), the NR/CDs inclusion complexation most likely occurs at the polar/apolar interfaces of mixed micelles. Under these conditions, the proximity of NR to AC (which is situated in the aqueous phase) is highest and the attenuation of the AC fluorescence peak is maximal. The effects observed at higher molar fractions of CDs require a consideration of the NP sizes obtained upon  $\gamma$ -CD-C<sub>10</sub>/NR/P80 self-assembly (Fig. 5).

### 3.3 Nanoparticle sizes

Quasi-elastic light scattering (QELS) was employed for determination of the particles sizes and distributions upon formation of mixed nanoassemblies with amphiphilic  $\gamma$ -CD-C<sub>10</sub>. Figure 5 shows that there is an increase in the size of the P80/NR assemblies upon addition of CDs. This result confirms that there is an interaction between the amphiphilic CDs and the P80/NR micelles. It also suggests a structural change in the NP organization at increasing CDs content. The formation of  $\gamma$ -CD-C<sub>10</sub>/NR/P80 assemblies leads to NP sizes in the range from 140 nm to 825 nm. Indeed, NR embedded in large NPs, different from micelles, cannot only be located at the polar/apolar interfaces. Its inclusion in the NPs interior will increase the distance to the AC fluorophore. Hence, the reduction of the AC fluorescence intensity should be smaller for bigger NPs.

## 4 Conclusions

The performed study demonstrated the inclusion complexation between novel biotransesterified amphiphilic cyclodextrins and a model drug (NR) in nanostructured environment created by multilevel self-assembly. A new FRET pair (AC donor/NR acceptor) was proposed for probing the location of the guest NR in P80 micelles or in the CDs nanocavities in NPs. Both the QELS and the fluorescence measurements were sensitive to the NP organization at varying  $\gamma$ -CD-C<sub>10</sub> content. The obtained results provided evidence for the occurrence of host-guest type of interactions in a nanostructured medium. The structural organization of the new NPs requires a separate investigation.

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